This article was downloaded by:

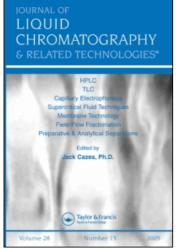
On: 24 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Journal of Liquid Chromatography & Related Technologies

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597273

Ion Exchange Application of Overpressured Thin-Layer Chromatography Huba Kalász^a; János Nagy^a

^a Department of Pharmacology, Semmelweis University of Medicine, Budapest, HUNGARY

To cite this Article Kalász, Huba and Nagy, János(1981) 'Ion Exchange Application of Overpressured Thin-Layer Chromatography', Journal of Liquid Chromatography & Related Technologies, 4: 6, 985 — 1005

To link to this Article: DOI: 10.1080/01483918108059600 URL: http://dx.doi.org/10.1080/01483918108059600

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

ION EXCHANGE APPLICATION OF OVERPRESSURED THIN-LAYER CHROMATOGRAPHY

Huba Kalasz and Janos Nagy

Department of Pharmacology, Semmelweis University of Medicine, H-1089 Budapest, Nagyvårad ter 4, HUNGARY.

ABSTRACT

The application of overpressured thin-layer chromatography introduced into the field of ion exchange chromatography. The basic differences between overpressured thin-layer chromatography and classical thin-layer chromatography are discussed including the distinction between the separations performed on thin-layer plates containing silica gel and a mixture of ion exchanger material and silica gel. The basic increase of flow velocity of solvent front with the aid of a pressurized ultra-micro chamber and the effect of flow velocity on the height equivalent of the theoretical plates are also presented. For basic amino acids, the flow velocity vs plate height curves show optima at a moderately high rate of development.

INTRODUCTION

Ion exchange application of overpressured thin-layer chromatography (OPTLC) is an extension of the progress in some distinct fields of thin-layer chromatography (TLC).

Several authors have used TLC on sorbent layer consisting of ion exchanger cellulose. These studies were expanded by Berger and Blanquest (1) as well as Lepri and coworkers (2) whe used mixtures of cellulose and ion exchanger resin. Devenyi and his coworkers (3-9) reported the construction and application of mechanically stable sorbent layer consisting of strongly acidic cation exchanger. These ready-made plates are supplied by Chinoin-Nagytėtėny (Budapest, Hungary) as Fixion 50 X 8 and Macherey-Nagel (Duren, FRG) as lonex 25. In his publications Dévenyi discussed the wide application of Fixion 50 X 8 in the quantitative and qualitative determination of amino acids from protein and peptide hydrolysates, body fluids, etc. The separation takes only few hours, the samples from the hydrolysates can be directly applied to the plates even from 6 N hydrochloric acid solution. The method can be used as a micro determination in the sequence analysis using endopeptidase (8-9). Ion exchange TLC seems to be a suitable method both before and in conjunction with automatic amino acid analysis.

On the other hand, theoretical and practical development, using OPTLC showed, that the vapor phase over the sorbent layer may be eliminated (10) and the

flow velocity of solvent front increased (11-12). These results were accomplished using silica gel TLC and organic solvents. For practical reasons, a major part of the illustrative experiments was performed with dye-substances (11-13).

Kaiser determined the basis of high performance thin-layer chromatography (HPTLC) and stated its advantages (14); namely, the short time of development and excellent resolution. HPTLC however needs not only the special HPTLC-chamber but also extra instrumentation for application of the sample and detection of the separated substances. The primary limit of HPTLC is not the apparatus requirement but the very limited distance of development.

Guiochon (15-16) explained the factors which determine TLC and HPTLC and stated that the present conditions of HPTLC can not ensure the optimal flow velocity over a certain distance of development.

Tyihak and coworkers (11-13, 17) developed the apparatus and the methodology of OPTLC including the pressurized ultra-micro chamber (PUM-chamber) for both circular (13) and linear development (11-12, 17).

These apparatus demonstrate the improvement of several hitherto unknown factors, these include:

a./ the role of vapor phase in TLC, the elimination of vapor phase over the sorbent layer and the
beneficial effect of its elimination on the reproducibility and uniformity of TLC experiments (11-12),

- b./ the possibility of constant velocity of solvent front during the development and over a short distance (11-12),
- c./ the possibility of increasing the rate of development (11-13, 17), i.e. the significant decrease of time of development,
- d./ the phenomenon, that the basic increase of flow velocity does not result in a decrease in the efficiency of TLC (17).

In this study the effect of vapor phase and solvent flow velocity on the characteristics of TLC and OPTLC using silica gel and Fixion 50 X 8 plates is compared.

APPARATUS

Set up for conventional TLC

The experiments were performed in Desaga Chambers.

The solvent system used were added 1 hour prior to development in each case. Saturation was achieved by lining the developing tank with filter paper, while

the non-saturated chamber was unlined with filter paper. The covered plates (very similar to Tyihak's UM-chamber (10)) serve to show the effect of the elimination of vapor phase over the sorbent layer when covered with a glass plate.

Set up for OPTLC

The sorbent layer is sandwiched between a plastic sheet and a foil which is tightly pressed onto the thin-layer by a gas pressure up to 300 kPa (about 3 atm). This gas pressure exists in the space between the membrane ant the upper support block. The upper support block is also the holder of the pressure inlet, pressure gauge, inlet for developing solvent system and 0-ring holding the membrane. The upper and lower support blocks are the frame of the whole set up and held together by several screw-like tools. The dimensions of the membrane are 200 x 200 mm; those of the support blocks are 230 x 230 mm.

EXPERIMENTS

Basic amino acids were separated on silica gel plates using propanol-ammonia, and chloroform-methanol-water as mobil phases. The $\mathbf{R_f}$ vs front distance data are given in Table 1. in the cases of:

Table 1.

The dependence of $R_{\mbox{\scriptsize f}}$ values of basic amino acids from the running distance of front. Silica gel sorbent layer and n. propanol - ammonia (1:1) were used.

front distance					R _e × 100	00			
(ww)	non-sa	turated	non-saturated chamber	satur	saturated chamber	amber	PUM-c	PUM-chamber	
	Arg	His	Lys	Arg	His	Lys	Arg	His	Lys
160	37	62	75	34	59	72	35	99	74
145	34	99	69	32	53	72	34	55	71
130	30	94	62	27	43	59	30	94	62
115	56	36	55	23	33	52	26	36	55
001	21	31	48	17	28	14	20	31	48
85	19	28	47	15	77	14	19	27	47
70	17	24	94	14	20	1 4	17	23	41
55	13	18	40	=	16	38	13	18	70
07	13	17	38	10	15	35	13	17	38

saturated chamber, non-covered sorbent layer (S - NC) non-saturated chamber, non-covered layer (NS - NC).

The front distance vs time diagrams show the situation in the cases of S-NC, NS-NC as well as S-C (saturated chamber, covered sorbent layer) and NS-C (non-saturated chamber, covered sorbent layer) developments with dichloro-methane and chlorform-methanol-water mobile phases. In addition to the chromatography in a conventional chamber (TLC), the experiments in the PUM-chamber are also shown using three different flow velocities. In a very similar arrangement, the same amino acids were also separated on Fixion 50 X 8 chromatoplates, using the saturated and non-saturated chambers as well as covered and non-covered sorbent; the front distance vs time diagrams are also presented (Figs. 1 and 2).

All plates were 200 mm x 200 mm in size.

TLC aluminium sheets silica gel 60 F₂₅₄, precoated were used (E. Merck, Darmstadt, FRG) and Fixion
50 X 8 chromatoplates (Chinoin-Nagyteteny, Budapest,
Hungary). The solvent systems for development of silica gel plates were:

- a./ n-propanol-25 % ammonia (1:1),
- b./ dichloromethane,

c./ chloroform-methanol-water (7:5:1),
the solvents for the Fixion 50 X 8 plates were:
d./ sodium citrate aqueous solutioon, pH = 4.3,
containing 1.2 normal sodium ion,
e./ sodium citrate aqueous solution, pH = 3.26,
containing 0.2 normal sodium ion.

The R_f values of amino acids investigated in the a./. c./, d./ and e./ solvent systems are given in Tables 1, 2, 3 and 4.

DISCUSSION

Fig. I shows the distance reached by the solvent front in silica gel precoated sheets in the case of S-NC, S-C, NS-C and NS-NC systems, using the conventional TLC. Fig. I also presents the results of experiments in PUM-chamber using different flow velocities. The strikingly common characteristics of the conventional TLC runs are, the further the solvent front, the slower its movement. At the same time, when OPTLC was used, a plot of front vs time gave nearly straight lines. The experiments in saturated and non-saturated chambers but with non-covered plates differ from each other significantly, while the two curves taken dur-

ing TLC of S-C and NS-C are essentially the same (Fig. 1). These curves demonstrate the basic role of vapor phase in the developing characteristics of conventional TLC. Also the fundamental role of the vapor phase is indicated since significant deviations were found with respect to the $R_{\rm f}$ values of amino acids (Tables 1 and 2).

Fig. 2 shows the solvent front distance vs time diagrams for Fixion 50 X 8 chromatoplates. For Fixion, the presence or absence of vapor phase plays only a minor role as both the running characteristics and the R_f values are practically independent from the saturation of the chamber (Fig. 2, Tables 3 and 4).

Fig. 3 shows the developing process of thin-layer chromatography in conventional— and in PUM-chamber. The series demonstrate a complete development in PUM-chamber while in the case of conventional TLC the solvent front moved much slower. In both cases, Fixion 50 X 8 plates were used, and the developing solution was citrate buffer at pH = 4.3 (d./).

Fig. 4 shows the plate height vs flow velocity of solvent front when basic amino acids are separated in PUM-chamber, on Fixion 50 X 8 plates and developed

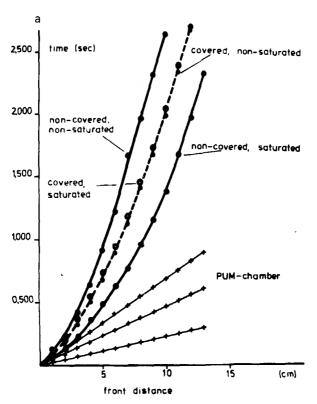
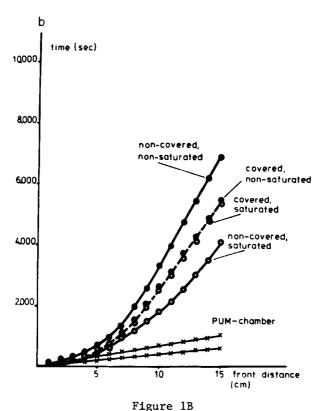


Figure 1. Diagrams are demonstrated when a volatile organic solvent (dichloromethane) was the running system (a.). The front distance vs. time of development data show difference in the case of NC-S and NC-NS chambers in solvent system chloroform-methanol-water (7:5:1) too, although the curves come close to each other (b.). Experiments were made on silica gel pre-coated layers.

in buffer d./. Each curve shows a definite maximum value, if the effect of flow velocity on the plate height is considered. Regarding the form of the curves, the ascending and the descending portions of the curves have to be considered separately. The descending part



J

shows the phenomenon, that the increase of the flow rate is possible up to a certain height of solvent front. This means an optimal flow rate of about 0.1 mm/sec in the case of the above mentioned chromatographic characteristics. At the same time, the quick ascent of the velocity vs plate height curves may be the consequence of a double effect, namely:

 $\label{eq:Table 2.} \textbf{R}_{\text{f}} \text{ values of amino acids. Running distance was 160 mm.}$

sorbent	silica gel						
solvent system	n. prop	anol-ammon (1:1)	1a	chlorofo	rm-methan (7:5:		
-chamber	S-NC	NS-NC	PUM-	S-NC	NS-NC	PUM-	
amino acid			R _f x	100		,	
Arg	34	37	35	00	00	00	
Lys	59	62	60	02	06	04	
His	72	75	74	08	12	10	
Phe	82	88	86	36	47	40	
Tyr	69	72	70	22	27	25	
DOPA	64	70	68	12	16	15	
Met	77	79	78	27	35	30	
Val	66	69	67	22	29	27	
Ala	65	67	66	11	15	13	
Gly	66	66	66	05	08	05	
Leu	73	80	74	33	41	35	
lle	71	76	75	31	39	34	
Thr	58	64	60	07	14	80	
Ser	50	62	57	05	08	06	
Pro	53	63	59	15	20	17	
HyPro	51	61	57	09	14	13	
Trp	70	74	71	29	41	35	
5-0H-Trp	71	72	71	17	22	20	
Glu	54	64	60	04	09	06	
Asp	49	62	55	03	06	04	

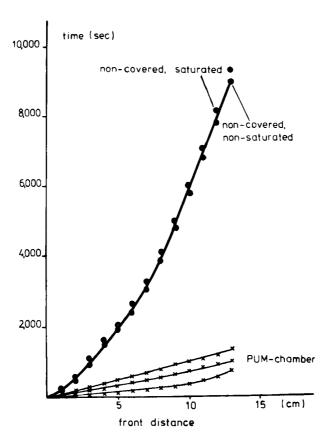


Figure 2. Fixion 50 X 8 chromatoplates were used in aqueous buffer and the front distance vs. time diagrams practically gave the same results in NC-S and NC-NS chambers.

- 1./ the increase of plate height is the result of the local nonequilibrium in consequence of the increase of flow velocity,
- 2./ a supplementary factor may be a slight overflow of the developing solvent, that is the pres-

Table 3.

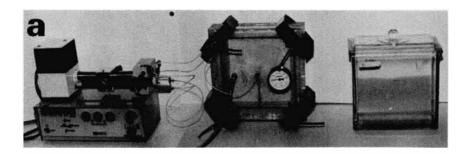
The dependence of $R_{
m r}$ of basic amino acids from the running distance of solvent front. Fixion 50 χ 8٦

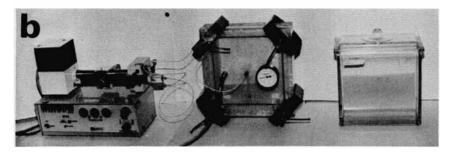
plates and citrate buffer at pH = 4.3 were used.	rate buffer at pH = 4	.3 were	.3 were used.) n : :) (
front distance (mm)				~	R _f × 100				
	non-sat	urated	non-saturated chamber	satur	saturated chamber	ambe r	PUM	PUM-chamber	
	Arg	His	Lys	Arg	His	Lys	Arg	His	Lys
160	16	25	41	16	23	07	16	25	41
145	91	25	141	91	23	04	91	25	41
130	16	25	41	91	23	04	91	25	4 3
115	16	25	14	91	23	04	91	25	۲۱
100	91	25	11	91	22	70	91	25	14
85	91	25	41	91	23	70	91	25	41
70	91	26	42	91	23	41	91	25	42
55	18	27	43	17	77	14	17	56	42
70	18	27	43	17	25	41	18	56	42

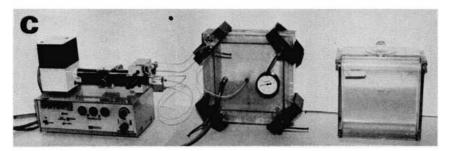
 ${\bf R_f}$ values of amino acids. Running distance was 160 mm.

Table 4.

sorbent	F	ixion 50 X	8 chromatoplates	
running system		buffer at .26 (e./)	citrate buffe pH = 4.3 (d.	r at /)
-chamber	NS-NC	PUM-	NS-NC	PUM-
amino acids		R _f	x 100	
Arg	01	01	16	16
Lys	05	05	41	41
His	03	03	25	25
Phe	11	11	35	35
Tyr	10	10	40	40
DOPA	16	16	52	52
Met	27	27	62	62
Val	40	40	70	71
Ala	46	45	76	80
Gly	50	50	74	77
Leu	17	17	58	59
He	23	23	61	61
Thr	65	66	83	83
Ser	61	61	80	80
Pro	44	45	64	64
Trp	03	03	14	14
5-OH-Trp	04	04	19	19
Asp	85	85	88	91
Glu	81	82	78	78







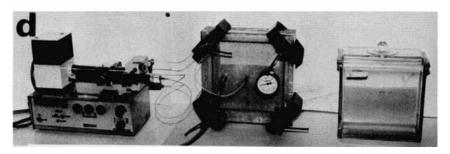
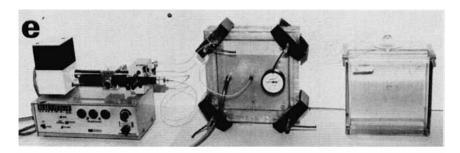


Figure 3(a-d)



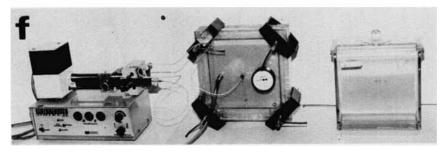


Figure 3. The processes of development are comparatively illustrated in conventional TLC- and in PUM-chambers (Figs. 3/a, 3/b, 3/c, 3/d, 3/e and 3/f). The start at the right beginning (3/a) and more stages of development are shown in PUM-chamber. The syringe-type pump (left), the PUM-chamber (middle) and the conventional TLC-chamber (right) are shown on each picture. Fixion 50 X 8 plates developed in sodium citrate buffer, pH = 4.2 were used.

sure on the foil is not sufficient to compensate for the pressure effect of the solvent supplied by the pump.

If only the first point is true, the optimal flow velocity can be stated by the above detailed way, and it is characteristic for the case of PUM-chamber, TLC

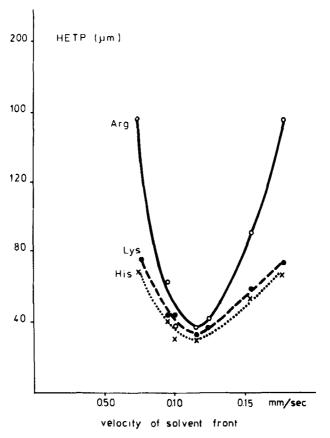


Figure 4. In case of Fixion 50 X 8 plates and PUM-chamber, the plate height vs. flow velocity curves show optima at a moderately high value of flow velocity of solvent front.

and OPTLC, when amino acids are separated on Fixion 50 X 8 chromatoplates. If the second point is fulfilled, the application of higher external pressure on the membrane will make possible smaller plate heights, i.e. better efficiencies and separations. To realize higher

external pressure (about 10 atm) on the membrane the basic reconstruction of the apparatus will be necessary, e.g. the change of plexy glass to steel, etc.

At the same time, these experiments suggest the possibility of both reaching very low plate height in the OPTLC experiments and the further decrease of the time necessary for the development.

The above results in the area of separation of amino acids by OPTLC indicate that

- a./ decrease of time of development, and
- b./ the finding of optimal flow velocity
 have been accomplished. These results allow the separation of amino acids in 10 18 minutes with good resolution instead of 100 400 minutes required by conventional TLC.

An additional point worth mentioning here is that the vapor phase does not play as major a role in ion exchange chromatography as it does when TLC silica gel layers are used.

REFERENCES

 Berger, J., Meyniel, G., Petit, J. et Blanquet, P., Eploy des resines échengeuses d'ions et de couches multiples succesives en chromatographie en couches minces., Bull. Soc. Chim. France 1963, 2662.

 Lepri, L., Desideri, P. G. and Coas, V., Chromatographic and Electrophoretic Behaviour of Primary Mono- and Diamines on Layers of Weak and Strong Ion Exchangers, J. Chromatog., 79, 129 (1973).

- Dévényi, T. and Zoltán, S., Thin-Layer Ion-Exchange Chromatography of Amino Acids, 7th International Symposium on Natural Products. 1970. Riga, U.S.S.R. p. 52.
- Dévényi, T., Thin-Layer Ion-Exchange Chromatography on Resin Coated Chromatoplates. I. Separation of Aromatic and Basic Amino Acids. Acta Biochim. Biophys. Acad. Sci. Hung. <u>5</u>, 435 (1970).
- Devenyi, T., Bati, J. and Fabian F., Detection and Determination of Tryptophan by Ion Exchange Chromatography, Acta Biochim. Biophys. Acad. Sci. Hung. 6, 133 (1971).
- Dévényi, T., Hazai, S., Ferenczi, J. and Báti, J., Thin-Layer Ion-Exchange Chromatography on Resin Coated Chromatoplates. V. One-Dimensional Separation of Amino Acids. Acta Biochim. Biophys. Acad. Sci. Hung. 6. 385 (1971).
- Dévényi, T., Báti, J., Kovács, J. and Kiss, P., Thin-Layer Ion-Exchange Chromatographic Screening Test for Aminoacidemias in Blood Samples Dried on Filter Paper. Acta Biochim. Biophys. Acad. Sci. Hung. 7. 237 (1972).
- 8. Sajgó, M. and Dèvènyi, T., Thin-Layer Ion-Exchange Chromatography on Resin-Coated Chromatoplates. VII. Rapid Determination of C-terminal Sequence on the Nanomol Scale. Acta Biochim. Biophys. Acad. Sci. Hung. 7. 233 (1972).
- Kisfaludy, L., Lbw, M. and Devenyi, T., Enzymatic Degradation of Peptides Containing Aminooxy Carboxylic Acids. Acta Biochim. Biophys. Acad. Sci. Hung. 6. 393 (1971).
- Tyihak, E. and Held, G., TLC in Pharmacognosy, in: Progress in Thin-Layer Chromatography and Related Methods. Vol. II., Niederwieser, A. and Pataki, G. eds., Ann. Arbor. Publ. Michigan, 1971. p. 183.
- Tyihak, E., Mincsovics, E. and Kalasz, H., New Planar Liquid Chromatographic Technique: Overpressured Thin-Layer Chromatography, J. Chromatog. 174, 75 (1979).

Downloaded At: 18:33 24 January 2011

- Mincsovics, E., Tyihak, E. and Kalasz, H., Resolution and Retention Behaviour of Some Dyes in Overpressured Thin-Layer Chromatography, J. Chromatog. 191. 293 (1980).
- Kalász, H., Nagy, J., Mincsovics, E. and Tyihák, E., Circular Development with Overpressured Thin-Layer Chromatography. J. Liquid Chromatography 3. 845 (1980).
- 14. Kaiser, R. E., The U-chamber. in: HPTLC High Performance Thin-Layer Chromatography. Zlatkis, A. and Kaiser, R. E. (eds.) Elsevier, Amsterdam, 1977.
- Guiochon, G. and Siouffi, A., Study of the Performance of Thin-Layer Chromatography. III. Flow Velocity of Mobile Phase. J. Chromatog. Sci. 16, 598 (1978)
- Guiochon, G. and Siouffi, A., Study of the Performance of Thin-Layer Chromatography. II. Band Broadening and Plate Height Equation. J. Chromatog. Sci. 16, 470 (1978).
- Kalasz, H., Tyihak, E. and Mincsovics, E., Overpressured (pressurized) Thin-Layer Chromatography. in: Recent Developments in Chromatography and Electrophoresis, 10. Frigerio, A. and McCamish, M. (eds.) in press.